

RESEARCH ARTICLE

Inflammatory Mediators in Tracheal Aspirates of Preterm Infants Participating in a Randomized Trial of Inhaled Nitric Oxide

Mandy Laube^{1*}, Elena Amann², Ulrike Uhlig³, Yang Yang³, Hans W. Fuchs⁴, Michael Zemlin^{5,6}, Jean-Christophe Mercier⁷, Rolf F. Maier⁵, Helmut D. Hummler², Stefan Uhlig³, Ulrich H. Thome¹

1 Center for Pediatric Research Leipzig, Hospital for Children & Adolescents, Division of Neonatology, University of Leipzig, Leipzig, Germany, **2** Division of Neonatology and Pediatric Critical Care, Department of Pediatrics, University of Ulm, Ulm, Germany, **3** Institute of Pharmacology and Toxicology, RWTH Aachen University, Aachen, Germany, **4** Department of Pediatrics, University Medical Center Freiburg, Freiburg, Germany, **5** Department of Pediatrics, University of Marburg, Marburg, Germany, **6** Department of Pediatrics, University of Saarland, Homburg, Germany, **7** Hôpital Universitaire Robert Debré, Paris, France

* mandy.laube@medizin.uni-leipzig.de



OPEN ACCESS

Citation: Laube M, Amann E, Uhlig U, Yang Y, Fuchs HW, Zemlin M, et al. (2017) Inflammatory Mediators in Tracheal Aspirates of Preterm Infants Participating in a Randomized Trial of Inhaled Nitric Oxide. PLoS ONE 12(1): e0169352. doi:10.1371/journal.pone.0169352

Editor: Lynn M Schnapp, Medical University of South Carolina, UNITED STATES

Received: May 27, 2016

Accepted: December 15, 2016

Published: January 3, 2017

Copyright: © 2017 Laube et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are presented in the manuscript and the materials described are freely available.

Funding: We acknowledge financial support by iNO Therapeutics. We declare that iNO Therapeutics had no role in the study design, data collection and analysis, their interpretation, in writing of the paper and in the decision to submit the publication. We further acknowledge support from the German Research Foundation (DFG) and Leipzig University within the program of Open Access Publishing. The

Abstract

Background

Ventilated preterm infants frequently develop bronchopulmonary dysplasia (BPD) which is associated with elevated inflammatory mediators in their tracheal aspirates (TA). In animal models of BPD, inhaled nitric oxide (iNO) has been shown to reduce lung inflammation, but data for human preterm infants is missing.

Methods

Within a European multicenter trial of NO inhalation for preterm infants to prevent BPD (EUNO), TA was collected to determine the effects of iNO on pulmonary inflammation. TA was collected from 43 premature infants randomly assigned to receive either iNO or placebo gas (birth weight 530–1230 g, median 800 g, gestational age 24 to 28 2/7 weeks, median 26 weeks). Interleukin (IL)-1 β , IL-6, IL-8, transforming growth factor (TGF)- β ₁, interferon γ -induced protein 10 (IP-10), macrophage inflammatory protein (MIP)-1 α , acid sphingomyelinase (ASM), neuropeptide Y and leukotriene B₄ were measured in serial TA samples from postnatal day 2 to 14. Furthermore, TA levels of nitrotyrosine and nitrite were determined under iNO therapy.

Results

The TA levels of IP-10, IL-6, IL-8, MIP-1 α , IL-1 β , ASM and albumin increased with advancing postnatal age in critically ill preterm infants, whereas nitrotyrosine TA levels declined in both, iNO-treated and placebo-treated infants. The iNO treatment generally increased nitrite TA levels, whereas nitrotyrosine TA levels were not affected by iNO treatment. Furthermore, iNO treatment transiently reduced early inflammatory and fibrotic markers associated with

funder had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

BPD development including TGF- β_1 , IP-10 and IL-8, but induced a delayed increase of ASM TA levels.

Conclusion

Treatment with iNO may have played a role in reducing several inflammatory and fibrotic mediators in TA of preterm infants compared to placebo-treated infants. However, survival without BPD was not affected in the main EUNO trial.

Trial registration

[NCT00551642](https://www.clinicaltrials.gov/ct2/show/study/NCT00551642)

Introduction

Survival of preterm infants is frequently associated with a chronic lung disease called broncho-pulmonary dysplasia (BPD). Infants who develop BPD often require long-term oxygen supplementation [1,2] and frequent re-admissions to hospitals [3,4], resulting in high health care costs [5]. Quality of life may be reduced, especially since pulmonary function abnormalities may persist into adulthood [6,7].

Lung injury induced by mechanical ventilation and oxygen supplementation triggers pro-inflammatory responses and repair processes [8–10]. Cytokines can be measured in tracheal aspirates (TA) and reflect the extent of inflammatory reactions [11,12]. It was reported that interleukin (IL)-1, IL-6, IL-8, intercellular adhesion molecule-1 (ICAM-1), macrophage inflammatory protein (MIP)-1 α , transforming growth factor (TGF)- β_1 and leukotriene B₄ (LTB₄) were increased within the first 10 days of life in the bronchoalveolar lavage (BAL) fluid of preterm infants who later developed BPD as compared to those who did not [13–19]. In a multicenter high frequency oscillatory ventilation trial [20], the outcome of the trial was predicted by IL-8 and LTB₄ TA levels [21]. One further important chemokine in lung injury that is critical for leukocyte activation is CXCL10 (IP-10) [22]. Another potentially interesting mediator is neuropeptide Y (NPY), because it may be involved in the so-called neuro-immune axis [23]. In addition, increased levels of glycolipids, such as ceramide, were found in the BAL fluid of patients with respiratory distress syndrome (RDS) [24] and in an ovine BPD model [25]. Furthermore, acid sphingomyelinase (ASM), an enzyme generating ceramide, was increased in ovine BPD [25] and in septic patients, where it correlated with their mortality [26]. These findings are remarkable, because the ASM pathway is critical for edema formation in many models of acute lung injury [27] including neonatal piglet models [28,29] and may also promote apoptosis in lung epithelial cells [30,31].

Nitric oxide (NO) is a gaseous mediator that—apart from its vasodilatory properties—has various effects on inflammation [32,33] and vascular endothelial growth factor (VEGF)-mediated tissue remodeling [34], both possibly modifying the development of BPD. In animal models of BPD, inhaled NO (iNO) has been shown to reduce lung inflammation, apoptosis and oxidative stress, to maintain surfactant activity, and to improve lung structure and alveolarization [33,35–41]. In humans, a randomized trial demonstrated that iNO reduced the incidence of BPD without increasing the risk of intracranial bleeding in mechanically ventilated preterm infants [42]. However, subsequent studies did not observe similar beneficial effects [43–45]. Aside from possible benefits, NO is a free radical, potentially capable of causing oxidative

tissue damage. Furthermore, it might combine with superoxide to form peroxynitrite, a powerful nitrating and tissue damaging substance [46]. Reactive oxygen and nitrogen species, such as peroxynitrite, also participate in lung injury [47,48]. Although they cannot be measured directly because of their instability, peroxynitrite formation can be estimated from the resulting nitration products, especially nitrated tyrosine residues on various proteins including surfactant protein A [49,50]. On the other hand, iNO may alleviate free radical toxicity, as NO has been shown to decrease lipid peroxidation and spare α -tocopherol by scavenging peroxy radicals [51,52].

Within a European multicenter trial of NO inhalation for preterm infants to prevent BPD (EUNO trial) [45], TA were collected at two study centers to determine the effects of iNO on pulmonary inflammation. We hypothesized that iNO reduces pro-inflammatory and pro-fibrotic cytokines and ASM in TA. Furthermore, we hypothesized that iNO may be associated with a higher amount of nitrite and nitrotyrosine possibly indicating increased peroxynitrite formation.

Materials and Methods

The EUNO trial

In brief, infants with a gestational age between 24 and 28 2/7 weeks were enrolled if they weighed at least 500 g and required surfactant or continuous positive airway pressure for RDS within 24 h of birth. Treatment was initiated within 2 h of enrollment, but not later than 26 h of life. Infants were randomized to receive either iNO (5 parts per million [ppm]) or placebo gas (nitrogen gas) for a minimum of 7 days and a maximum of 21 days in a double-blind fashion [45]. BPD at 36 weeks' postmenstrual age was defined by the physiological criteria of Walsh and colleagues [53,54]. Infants enrolled at two of the study centers (Marburg and Ulm, Germany) were eligible for TA sampling if they were endotracheally intubated for mechanical ventilation.

Tracheal aspirate sampling

This sub-study was specifically approved by the institutional review board (INOT27, reference number 220/2004, Ulm, Germany) which also oversaw the main trial. Written informed parental permission was obtained specifically for the sub-study (collection of TA samples), in addition to permission for enrollment in the study. After initiation of iNO or placebo gas treatment TA was collected during normal medically indicated endotracheal suctioning procedures on post-natal days (PD) 2, 4, 7, 14 and 21, unless the infant was extubated earlier. By protocol, infants also received the randomized gas at least until they were weaned off of ventilatory support or reached PD21. Herein, we report the TA data obtained between PD2 and PD14, since increasing numbers of infants were extubated within the study period resulting in a low statistical power for data obtained on PD21. If less than 4 specimens per sampling day were obtained, further specimen were collected on the following day. No samples were collected within 4 hours of a surfactant instillation. A standardized procedure was used. For sampling, a sterile mucus trap was inserted in the suctioning system, and 1 ml/kg birth weight of normal saline was instilled into the endotracheal tube, and the ventilator briefly reconnected (3–5 breaths). The suction catheter was then flushed with 0.5 ml normal saline. TA was transferred to an appropriate tube and immediately centrifuged with 140 x g at 4°C for 10 minutes. The supernatant was transferred to cryotubes and frozen immediately below -20°C. Within 72 hours, cryotubes were transferred to a -80°C freezer and held at -80°C until ready for shipment to the laboratory, which was done on dry ice.

Tracheal aspirate analyses

All analyses were performed at the Institute of Pharmacology and Toxicology of the RWTH Aachen (Aachen, Germany). For the parameters IL-1 β , IL-6, IL-8, IP-10 and MIP-1 α a Bio-Plex Cytokine assay (Bio-Rad Laboratories, Munich Germany) was used [55]. Enzyme-linked immunosorbent assays (ELISA) were used for albumin (# EA2201-1; AssayPro; St. Charles, USA), nitrotyrosine (# HK501; Cell Sciences; Canton, USA), and TGF- β ₁ (# DB100B; R&D Systems GmbH, Wiesbaden-Nordenstadt, Germany). Furthermore, competitive binding assays were employed for LTB₄ (# KGE006B; R&D Systems GmbH) and NPY (# EK-049-03; Phoenix Europe GmbH, Karlsruhe, Germany). Nitrite concentration was analyzed using a Griess reaction assay (# KGE001; R&D Systems GmbH). All assays were performed according to the manufacturer's recommendations. For ASM, a proprietary assay was used [56]. Samples from the same patient and day were pooled to increase the amount and decrease variations of dilution. No attempt to normalize data was made since no uniformly accepted standard is currently available. We expressed the data per milliliter of TA as recommended by the European Respiratory Task Force on Bronchoalveolar Lavage in children [57,58] and reported by others [59,60].

Statistical analyses

Demographic and clinical outcome data were compared between iNO- and placebo-treated groups by Mann Whitney U test or Fisher's Exact test as appropriate using GraphPad Prism (version 6.05; GraphPad Software, Inc, San Diego, CA, USA). Measured TA concentrations were compared by mixed model two-way (factors being time and treatment) analyses of variance (ANOVA) with a heterogeneous first-order autoregressive covariance structure using SAS software 9.4 (GLIMMIX procedure, SAS Institute, Cary, NC). Post-tests were performed for the treatment effect on each day and *p*-values were adjusted for multiple comparisons by the simulated Shaffer procedure. In the figures, the effect of time is denoted below the x-axis, the overall effect of treatment to the right of the graphs, and the treatment effects on each single day at the respective time points.

Results

During the recruitment period from the years 2006–2008, 67 infants were enrolled in the original EUNO trial in the two study centers. Of these 67 infants, 49 infants were recruited for this sub-study. No TA could be obtained from 6 infants, because of extubation within the first 24 h of life in 4 infants and missing parental consent for TA sampling in 2 infants, resulting in 43 infants available for TA sampling. Of these 43 infants, 25 received iNO and 18 placebo gas. Demographic data were similar between the iNO- and placebo treated groups (Table 1). Clinical sepsis was defined by the maximal C-reactive protein (CrP) exceeding 20 mg/L within the first 72 h of life. Blood cultures were negative for all infants. Mean airway pressures and FiO₂ represent maximum values observed before initiation of iNO or placebo gas treatment.

TA levels were compared by mixed model ANOVA (Table 2) determining the effect of iNO treatment and postnatal age on the analyzed variables. Furthermore, the effect of iNO treatment was analyzed for each measured time point from PD2 to PD14. Nitrite levels in TA samples of iNO-treated infants were significantly higher over the complete study period ($p < 0.01$; Fig 1A). In particular, nitrite TA levels were significantly elevated by iNO on PD2 (iNO: $4.26 \pm 4.52 \mu\text{M}$ [mean \pm SD] versus control: $1.14 \pm 1.18 \mu\text{M}$; $p < 0.05$), PD4 (iNO: $2.66 \pm 1.87 \mu\text{M}$ versus control: $1.0 \pm 0.79 \mu\text{M}$; $p < 0.01$) and PD14 (iNO: $3.16 \pm 1.54 \mu\text{M}$ versus control: $1.39 \pm 2.46 \mu\text{M}$; $p < 0.05$). Postnatal age did not affect nitrite TA levels which were constant over the study period. In contrast, nitrotyrosine TA concentrations were not different between iNO-treated infants and the

Table 1. Demographic characteristics of the infants.

	iNO	Control	p-values
Number of patients	25	18	
Gestational age (weeks)*	26 (24 1/7-28 2/7)	26 1/7 (24–28 2/7)	0.812
Birth weight (g)*	800 (530–990)	815 (630–1230)	0.142
Male	14 (56.0%)	11 (61.1%)	0.765
Race (white)	25 (100%)	18 (100%)	1.000
Prenatal steroids	24 (96%)	18 (100%)	1.000
Apgar score 5-min*	8 (2–10)	9 (3–10)	0.633
Apgar score 10-min*	9 (7–10)	10 (5–10)	0.400
Surfactant replacement therapy	20 (80%)	16 (88%)	0.680
Death	1 (4%)	1 (6%)	1.000
BPD	5 (20.8%)	2 (11.8%)	0.679
Duration of ventilation (days)*	47 (9–141)	39 (17–82)	0.209
Therapy duration (days)*	21 (8–22)	21 (11–22)	0.605
Maternal chorioamnionitis	11 (44.0%)	11 (64.7%)	0.223
PPROM	5 (20.0%)	5 (29.4%)	0.717
Duration of PPRM (h)*	133 (23–600)	148 (38–878)	0.841
Mean airway pressures*	7.5 (0.8–26)	9 (5–28)	0.081
Sepsis	3 (12%)	5 (29.4%)	0.247
FiO ₂ *	0.6 (0.31–1)	0.6 (0.28–1)	0.820

*Median (minimum-maximum), Mann Whitney U test; all others: Fisher's exact test.
 PPRM: preterm prolonged rupture of membrane; FiO₂: fraction of inspired oxygen.

doi:10.1371/journal.pone.0169352.t001

placebo-treated control group, whereas nitrotyrosine levels decreased significantly over the study period in both groups ($p < 0.05$; Fig 1B). On PD7 and PD14 the majority of TA samples were negative for nitrotyrosine in both groups.

TGF- β_1 TA concentrations were significantly decreased by iNO on PD2 (iNO: 193 ± 116 pg/ml versus control: 466 ± 353 pg/ml; $p < 0.05$; Fig 2A). Furthermore, TGF- β_1 levels were

Table 2. Mixed model ANOVA (p-values).

	iNO treatment	Postnatal age
Nitrotyrosine	$p = 0.6468$	$p = 0.0312$
Nitrite	$p = 0.0028$	$p = 0.9801$
ASM	$p = 0.1117$	$p < 0.0001$
TGF- β_1	$p = 0.0224$	$p = 0.0575$
IP-10	$p = 0.3126$	$p < 0.0001$
IL-1 β	$p = 0.8465$	$p < 0.0001$
IL-8	$p = 0.352$	$p = 0.0202$
IL-6	$p = 0.9856$	$p = 0.0407$
MIP-1 α	$p = 0.7646$	$p < 0.0001$
Albumin	$p = 0.9069$	$p = 0.0073$
LTB ₄	$p = 0.9153$	$p = 0.3528$
NPY	$p = 0.5156$	$p = 0.8368$

P-values of the main factor of the two-way ANOVA: treatment and time (postnatal age). None of the interaction effects was significant.

doi:10.1371/journal.pone.0169352.t002

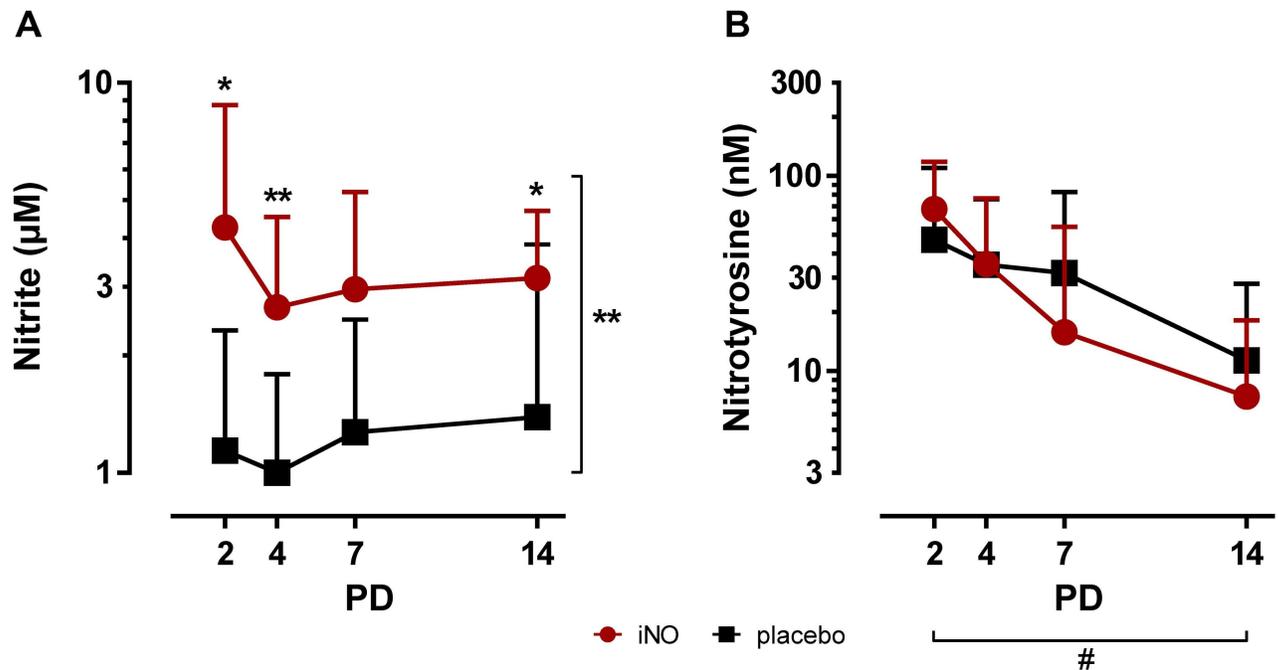


Fig 1. Nitrite and nitrotyrosine TA concentrations in infants treated with iNO compared to placebo-treated controls. Serial TA samples were obtained from PD2 to PD14. Data are displayed as the mean of TA levels and SD on a logarithmic scale. **A:** Nitrite TA levels were overall increased by iNO treatment (**, $p < 0.01$) and specifically elevated on PD2 (*, $p < 0.05$), PD4 (**, $p < 0.01$) and PD14 (*, $p < 0.05$) in iNO-treated infants. **B:** Nitrotyrosine TA levels were not altered by iNO treatment and decreased with increasing postnatal age of the infants from PD2 to PD14 (#, $p < 0.05$). PD: postnatal day.

doi:10.1371/journal.pone.0169352.g001

lower in iNO-treated infants over the entire study period ($p < 0.05$; Fig 2A). Postnatal age did not significantly affect TGF- β_1 TA levels. IL-1 β TA levels were not significantly affected by iNO treatment (Fig 2B). However, postnatal age strongly affected IL-1 β TA levels, which significantly increased from PD2 to PD14 in both groups ($p < 0.001$; Fig 2B). IL-6 TA concentrations were not affected by iNO treatment, but significantly increased with advancing postnatal age in both groups ($p < 0.05$; Fig 2C). NPY levels, in contrast were altered neither by postnatal age nor by iNO treatment (Fig 2D).

Among the chemokines, IP-10 TA levels were significantly decreased by iNO treatment on PD2 (iNO: $892 \pm 1,028$ pg/ml versus control: $1,940 \pm 1,501$ pg/ml; $p < 0.01$; Fig 3A). On PD4, PD7 and PD14 no differences for IP-10 were observed between the study groups. Postnatal age strongly affected IP-10 TA levels which significantly increased from PD2 to PD14 ($p < 0.001$). In addition to TGF- β_1 and IP-10, IL-8 TA concentrations were significantly reduced by iNO on PD2 with $3,485 \pm 2,140$ pg/ml in the iNO group compared to $14,262 \pm 22,931$ pg/ml in the placebo-treated control group ($p < 0.05$; Fig 3B). No difference for IL-8 was observed on PD4, PD7 and PD14 between iNO-treated infants and the placebo-treated control group. Furthermore, IL-8 TA levels significantly increased from PD2 to PD14 in both groups ($p < 0.05$). MIP-1 α TA levels were not significantly affected by iNO, but strongly increased in both, iNO- and placebo-treated infants from PD2 to PD14 ($p < 0.001$, Fig 3C). In contrast, LTB $_4$ TA levels were neither affected by postnatal age nor by iNO treatment (Fig 3D).

ASM TA concentrations were similar between iNO- and placebo-treated infants on PD2, PD4 and PD7, but the iNO group showed elevated ASM activities on PD14 with $11,631 \pm 6,822$ pM/mg/h compared to the placebo-treated control group with $5,589 \pm 1,713$ pM/mg/h ($p < 0.05$; Fig 4A). Moreover, ASM TA levels strongly increased over the study

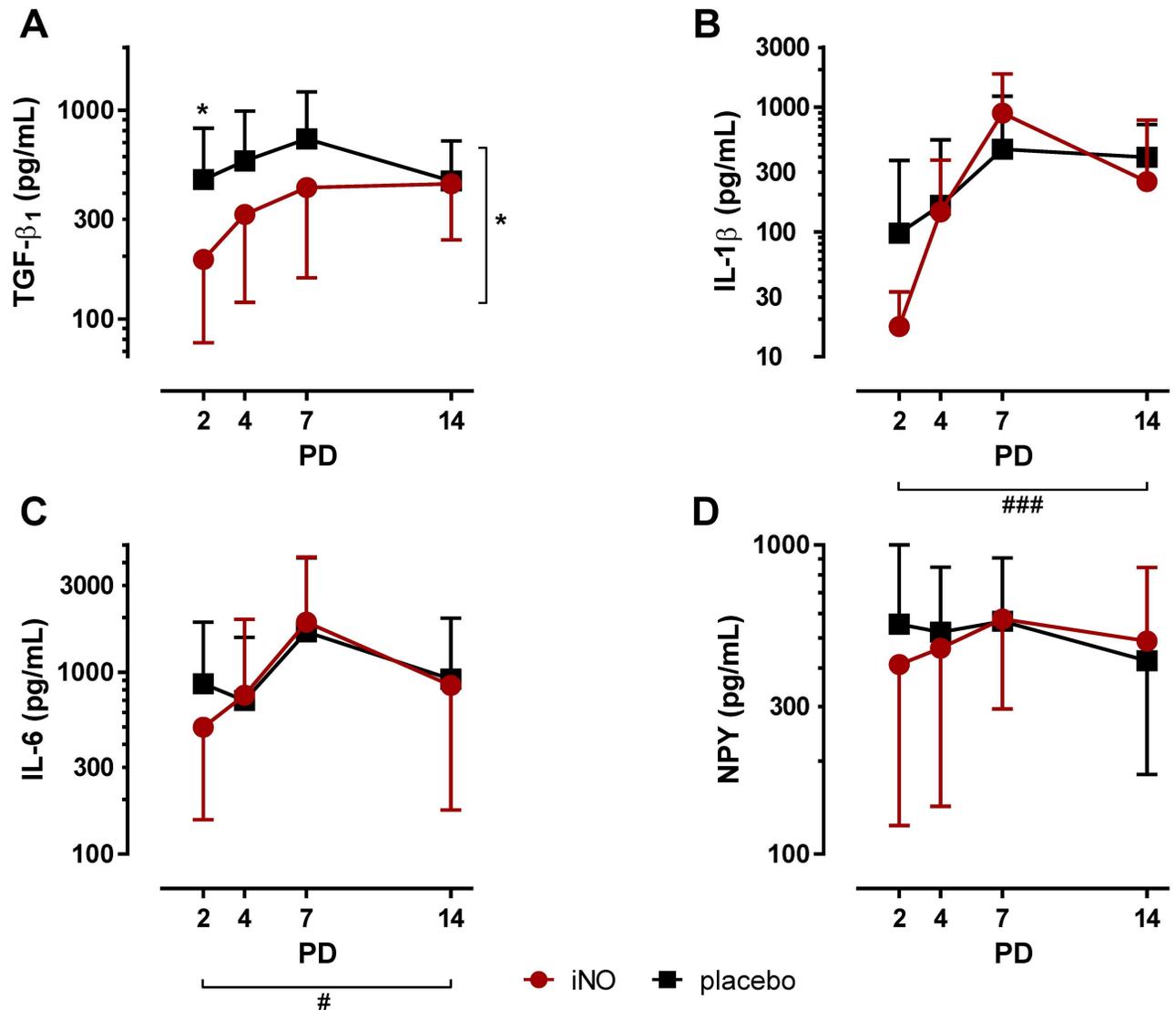


Fig 2. Cytokine TA concentrations in infants treated with iNO compared to placebo-treated controls. Serial TA samples were obtained from PD2 to PD14. Data are displayed as the mean of TA levels and SD on a logarithmic scale. **A:** TGF-β₁ TA levels were overall decreased by iNO treatment (*, p<0.05) and specifically decreased on PD2 (*, p<0.05) in iNO-treated infants. **B:** IL-1β TA levels were not significantly affected in iNO-treated infants and increased with advancing postnatal age from PD2 to PD14 in both groups (###, p<0.001). **C:** IL-6 TA levels were not altered by iNO treatment and increased with advancing postnatal age from PD2 to PD14 in both groups (#, p<0.05). **D:** NPY TA levels were not altered by iNO treatment or postnatal age. PD: postnatal day.

doi:10.1371/journal.pone.0169352.g002

period from PD2 to PD14 in both groups (p<0.001). Finally, albumin TA levels were not affected by iNO treatment as no significant differences were observed between the study groups at PD2 to PD14, but advancing postnatal age significantly elevated albumin TA levels over the study period (p<0.01; Fig 4B).

In conclusion, iNO treatment increased the nitrite and ASM concentrations in TA samples of preterm infants at individual time points, but had no effect on nitrotyrosine TA levels. Furthermore, TGF-β₁ TA levels were generally lower in iNO-treated infants, and IP-10 and IL-8 TA concentrations were transiently reduced by iNO treatment on PD2. Postnatal age, affected almost every analyzed TA variable, demonstrated by significant effects on nitrotyrosine, ASM, IP-10, IL-1β, IL-8, IL-6, albumin and MIP-1α concentrations.

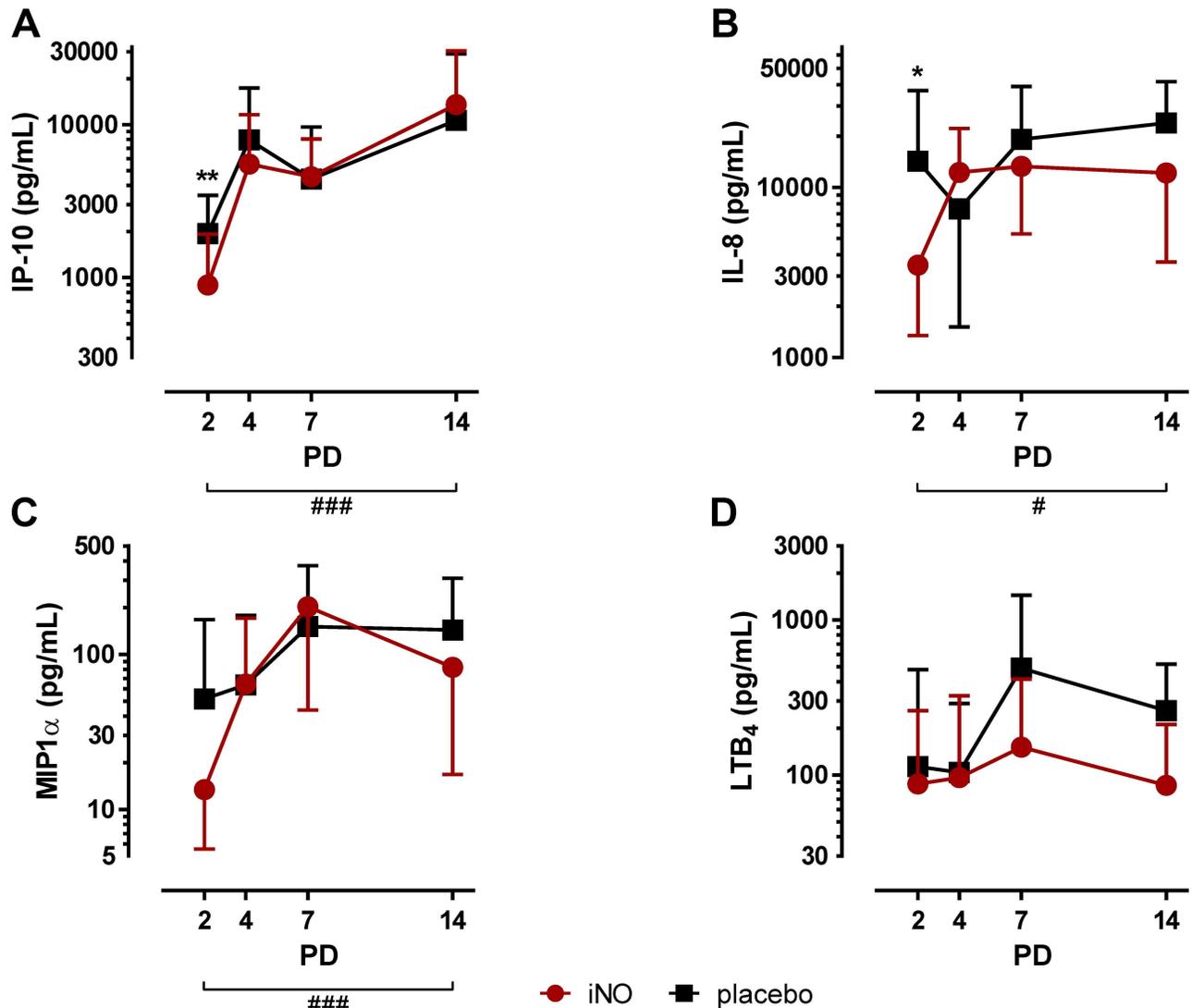


Fig 3. Chemokine TA concentrations in infants treated with iNO compared to placebo-treated controls. Serial TA samples were obtained from PD2 to PD14. Data are displayed as the mean of TA levels and SD on a logarithmic scale. **A:** IP-10 TA levels were significantly decreased on PD2 in iNO-treated infants (**, $p < 0.01$) and increased with advancing postnatal age from PD2 to PD14 in both groups (###, $p < 0.001$). **B:** IL-8 TA levels were significantly decreased on PD2 in iNO-treated infants (*, $p < 0.05$) and increased with advancing postnatal age from PD2 to PD14 in both groups (#, $p < 0.05$). **C:** MIP-1 α TA levels were not affected by iNO treatment and increased with advancing postnatal age from PD2 to PD14 in both groups (###, $p < 0.001$). **D:** LTB₄ TA levels were not altered by iNO treatment or postnatal age. PD: postnatal day.

doi:10.1371/journal.pone.0169352.g003

Discussion

Inflammation is crucially involved in the development of BPD and results from an imbalance between pro- and anti-inflammatory mediators [61]. Mechanical ventilation triggers the pulmonary influx of neutrophils and macrophages that produce a variety of cytokines and other signalling molecules [61]. Herein, we sought to determine the effect of iNO treatment on pulmonary inflammatory mediators in TA of preterm infants as part of the EUNO trial [45]. Our study showed that iNO treatment of preterm infants significantly reduced early TA levels of TGF- β_1 , IP-10 and IL-8. While the effect of iNO on TGF- β_1 persisted throughout the study, the effects on IP-10 and IL-8 were transient only. Furthermore, nitrite TA levels were

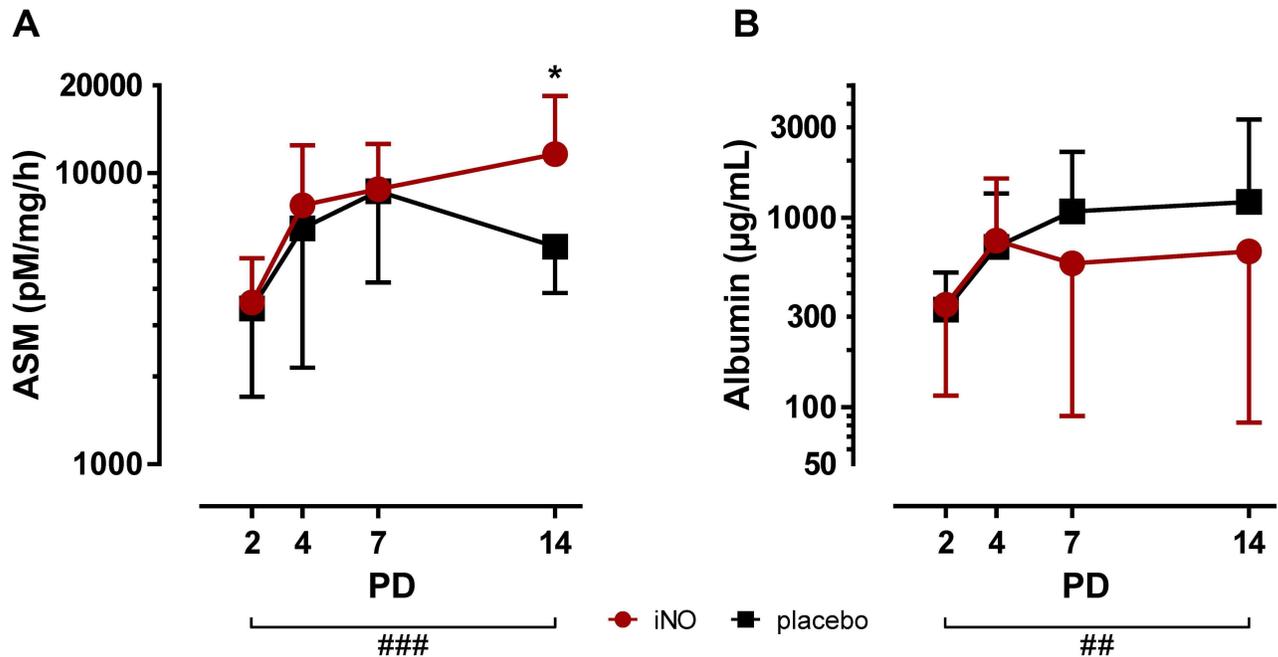


Fig 4. ASM and albumin TA concentrations in infants treated with iNO compared to placebo-treated controls. Serial TA samples were obtained from PD2 to PD14. Data are displayed as the mean of TA levels and SD on a logarithmic scale **A**: ASM TA levels were significantly elevated on PD14 in iNO-treated infants (*, $p < 0.05$) and increased with advancing postnatal age from PD2 to PD14 in both groups (###, $p < 0.001$). **B**: Albumin TA levels were not altered by iNO treatment and increased with advancing postnatal age from PD2 to PD14 in both groups (##, $p < 0.01$). PD: postnatal day.

doi:10.1371/journal.pone.0169352.g004

increased by iNO treatment throughout the study period and ASM levels were increased after 2 weeks. To our knowledge, only one study has analyzed the impact of iNO treatment on pulmonary inflammatory mediators in preterm infants. In contrast to our results, this study found no significant changes in TA concentrations of IL-8, IL-1 β or TGF- β ₁ induced by iNO [62]. These discrepancies might be due to the different study design, since we started iNO treatment within the first 24 h of life, employing low-dose iNO, whereas the study of Truog and colleagues started iNO treatment at PD7 with 20 ppm iNO. Most iNO effects observed in our study occurred early and transiently at PD2 and might not be detectable once the inflammatory response is already established.

NO is rapidly oxidized *in vivo* and the concentration of its metabolites, mainly nitrite and nitrate serve as biomarkers for NO [63]. Hence, iNO has been shown to dose-dependently affect nitrite and nitrate concentrations in TA and plasma, thereby demonstrating an effective delivery of iNO to the lung and the systemic circulation [64]. In line with this, we observed elevated TA levels of nitrite at PD2 and PD4, which remained elevated until PD14. In the presence of superoxide, NO may form peroxynitrite, which nitrates the phenolic residues of tyrosine, forming nitrotyrosine. We did not observe an increased nitrotyrosine TA concentration in the iNO group, and nitrotyrosine levels similarly decreased in both groups during the study period, which may be related to the diminishing exposure to supplemental oxygen with increasing postnatal age. A previous study showed that plasma nitrotyrosine concentrations were elevated during the first month of life in infants who developed BPD [65]. Another study on the therapeutic effects of iNO demonstrated that preterm infants whose nitrotyrosine levels decreased within the first 72 h of life were more likely to wean off of mechanical ventilation [66]. In agreement, another trial using iNO for preterm infants did not detect any changes of plasma nitrotyrosine

levels induced by iNO, nor was the nitrotyrosine concentration altered in infants with BPD compared to infants without BPD [67,68].

Early increases of pro-fibrotic TGF- β_1 in TA have been demonstrated in preterm infants that subsequently developed BPD [14], which was most pronounced on PD2 to PD4 [14,60]. Therefore, an increase of TGF- β_1 may represent an early event in the process leading to BPD that precedes abnormalities in lung function due to tissue remodeling and fibrosis (see review [69]). We demonstrated significantly lower TGF- β_1 TA levels in iNO-treated infants from PD2 on. Most important for the preterm lung, TGF- β_1 has been shown to inhibit epithelial cell maturation and the synthesis of phospholipids and surfactant proteins A, B and C in human fetal lung explants [70]. In agreement, one study demonstrated a transiently improved surfactant function in preterm infants undergoing iNO treatment [71]. TGF- β_1 activates fibroblast differentiation into myofibroblasts [72] and induces alveolar and bronchial epithelial cells to undergo epithelial-mesenchymal-transition [73,74], which contributes to interstitial thickening and fibrosis *in vivo* [75]. Notably, exogenous NO has been shown to attenuate epithelial-mesenchymal-transition induced by TGF- β_1 in alveolar epithelial cells [76] and iNO therapy has been shown to reduce hyperoxia-induced fibrin deposition and septal thickening in rat pups [77]. Interestingly, TGF- β_1 has recently been shown to inhibit β_2 -adrenergic receptor-mediated fluid transport across rat alveolar type II cell monolayers and alveolar fluid clearance (AFC) *in vivo* by down-regulation of β_2 -adrenergic receptors [78,79]. AFC is crucially involved in resolution of pulmonary fluid at birth and a TGF- β_1 -mediated AFC reduction possibly contributes to prolonged ventilator dependence of preterm infants leading to the development of BPD. Therefore, a reduction of early TGF- β_1 levels in preterm infants at risk of developing BPD by iNO treatment might suggest a beneficial clinical outcome.

Our study further showed that iNO treatment reduced the TA levels of different pro-inflammatory cytokines. First, the downstream target of IFN- γ , the chemokine IP-10 showed increasing TA concentrations during the study period in both groups, yet iNO treatment significantly reduced IP-10 levels at PD2, compared to the placebo-treated group. In a study of ventilated preterm infants, the IFN- γ and IP-10 TA levels were higher within the first 48 h of life in infants that developed BPD or died [80,81], suggesting that increases in IFN- γ and IP-10 levels precede neutrophil infiltration and could therefore represent critical early response molecules in the development of BPD [82], similar to what has been suggested for ARDS [22]. Thus, the early reduction of IP-10 levels by iNO might indicate a diminished inflammatory status in those infants that might be accompanied by a reduced recruitment of inflammatory cells to the lung. This is supported by an experimental BPD model of mechanically ventilated premature lambs in which low-dose iNO (5 ppm) decreased early lung neutrophil recruitment and accumulation [33]. However, the effect of iNO on IP-10 levels was transient. In addition, increased IL-8 TA levels have been described on PD1 and PD3 in preterm infants who subsequently developed BPD, preceding the neutrophil infiltration [83–85]; however, elevated TA IL-8 levels also seem to be associated with BPD development in older infants [13,86]. Here we found a rise of IL-8 TA concentrations during the study period and iNO treatment significantly reduced IL-8 TA levels at PD2. Since early IL-8 TA elevations supposedly precede neutrophil infiltrations in BPD risk infants [83], an early reduction induced by iNO might reduce neutrophil recruitment to the lung. In agreement with our results, iNO treatment has been shown to reduce BAL IL-8 concentrations and neutrophil infiltration in a pig model of lung injury [87] and human patients with ARDS [88]. Similar to IP-10, the effect of iNO on IL-8 levels was transient. The reason for this observation is currently unknown, but decreasing NO absorption or an elevated first-pass effect seems unlikely, since nitrite TA levels were relatively stable over the study period. Because infants are extubated as soon as possible, thus limiting TA sampling, a selection towards the more serious cases or patients in which iNO treatment failed to improve inflammatory mediator levels

possibly contributes to the early transient effects of iNO. Another possible reason for the transient effect of iNO might be due to the development of tolerance to the clinical and physiological effects of NO upon continuous administration [89]. The mechanism of tolerance to NO donors has been thoroughly investigated, especially in the treatment of cardiovascular diseases, but remains mainly elusive and highly debated. Notably, tolerance to nitrovasodilators generally begins to develop within 24 to 48 h of continuous application [89], and could thus be an explanation for the early transient effects of iNO observed in our study. Next-generation NO donors with improved pharmacokinetic properties might be able to prolong the anti-inflammatory effects of NO treatment in critically ill preterm infants.

We further observed increased ASM TA activities in iNO-treated infants on PD14. Although the function of circulating ASM is presently unknown, elevated ASM levels were found in different respiratory disorders (see reviews [27,90]), and we demonstrated increasing ASM activities throughout the current study in critically ill infants of both groups. Currently, it is unknown if BPD development in preterm infants is associated with ASM activity or ceramide levels. Many agonists, including tumor necrosis factor (TNF)- α and platelet activating factor (PAF) stimulate ceramide production [90] and PAF-induced pulmonary edema is partly mediated by ASM and ceramide [56]. On the other hand, inhibition of the ASM pathway as well as the *de novo* synthesis pathway of ceramide was shown to elevate IL-8 production in TNF- α stimulated respiratory epithelial cells, suggesting that ASM and ceramide may be involved in terminating an ongoing IL-8 production [91]. By contrast, ceramide has been shown to trigger IL-1 β release in a mouse model of acute lung injury [92]. Yet, IL-1 β TA levels were not elevated by iNO at PD14, suggesting that the up-regulation of ASM TA activities did not elevate the level of inflammatory mediators analyzed in our study. Finally, IL-6, albumin, LTB₄ and NPY TA levels were not significantly altered by iNO treatment compared to placebo-treated infants, but in contrast to LTB₄ and NPY, IL-6 and albumin TA levels increased throughout the study in both groups.

Several clinical trials have evaluated whether iNO reduces the mortality or the incidence of BPD in preterm infants with sometimes contradictory results [42,67]. The EUNO trial demonstrated that the early use of low-dose iNO (5 ppm) in very premature infants did not improve survival without BPD [45]. Inconsistencies between different iNO trials may be due to differences in dose, duration, age at treatment initiation, study population and/or other factors [93]. Therefore, results of these clinical trials are not conclusive and the use of iNO treatment for preterm infants to prevent BPD is currently not recommended [94]. Notably, a multicenter, randomized trial comparing high-frequency oscillatory ventilation with conventional ventilation in the early treatment of respiratory disease in very preterm infants showed no difference for the primary endpoint death or BPD incidence [95]. However, the follow-up of this trial demonstrated superior lung function at 11 to 14 years of age in former infants assigned to high-frequency oscillatory ventilation [96]. This suggests a beneficial outcome, although BPD incidence did not differ in the initial trial. With regard to this long term outcome, follow-up results for iNO therapy are currently awaited and BPD incidence possibly constitutes an imprecise endpoint to predict clinical benefits.

A limitation of this study was the small sample size, increasing the likelihood of statistical errors. Moreover, because in clinical practice infants are extubated as soon as feasible to limit potential damage from continuing ventilation, the number of infants of whom TA could be obtained declined with increasing postnatal age. This results in fewer TA samples and hence lower statistical power for TA analyses of infants older than one week. Furthermore, because of the limited sample size not all mediators could be measured from each TA sample. Therefore, subsequent trials with larger groups are required to confirm these results. We further did not include clinical characteristics such as maternal chorioamnionitis, PPRM, ventilator parameters, FiO₂ requirements or sepsis as confounding variables

in our analyses, which may play a role in release of inflammatory mediators. In addition, it is controversial whether TA should be corrected for dilution, using techniques such as albumin content, urea or secretory immunoglobulin A concentrations. Currently, no uniformly accepted correction factor to normalize cytokine TA levels exists, which impedes comparison of results obtained by different normalization procedures. As recommended by the European Respiratory Task Force on Bronchoalveolar Lavage in children [57,58], we did not correct our results for the dilution during TA sampling, and expressed our data per milliliter of TA.

Conclusions

In conclusion, the study showed that the TA levels of IP-10, IL-6, IL-8, MIP-1 α , IL-1 β , ASM and albumin increased with advancing postnatal age in critically ill preterm infants, whereas nitrotyrosine TA levels declined in both iNO-treated and placebo-treated infants. Furthermore, iNO treatment strongly increased nitrite TA levels throughout the study period. Besides, a delayed increase of ASM TA levels induced by iNO was detected. A beneficial effect of iNO was demonstrated on early inflammatory/fibrotic markers including TGF- β ₁, IP-10 and IL-8. Whether the demonstrated reduction of early inflammatory mediators possibly improves long-term lung function has to be determined in follow-up studies, since BPD incidence was not affected by iNO treatment in the EUNO trial [45].

Acknowledgments

The authors wish to thank iNO Therapeutics for financial support. We want to thank Nadine Ruske for excellent technical assistance. Furthermore, we thank the staff of the Ulm and Marburg NICU for collecting and centrifuging samples, and infants and their parents for participating.

Author Contributions

Conceptualization: UHT HDH SU J-CM.

Data curation: ML EA.

Formal analysis: SU.

Funding acquisition: UHT HDH J-CM.

Investigation: EA UU YY SU.

Methodology: UHT SU.

Project administration: UHT HDH.

Resources: HWF MZ RFM UU SU UHT HDH.

Supervision: UHT.

Validation: UU.

Visualization: ML SU.

Writing – original draft: ML UHT SU.

Writing – review & editing: ML EA UU YY HWF MZ J-CM RFM HDH SU UHT.

References

1. Monin P, Vert P. The management of bronchopulmonary dysplasia. *Clin Perinatol*. 1987; 14: 531–49. PMID: [3311538](#)
2. Nickerson BG. Bronchopulmonary dysplasia. Chronic pulmonary disease following neonatal respiratory failure. *Chest*. 1985; 87: 528–35. PMID: [3884289](#)
3. Katz R, McWilliams B. Bronchopulmonary dysplasia in the pediatric intensive care unit. *Crit Care Clin*. 1988; 4: 755–87. PMID: [3052708](#)
4. Horst PS. Bronchiolitis. *Am Fam Physician*. 1994; 49: 1449–53, 1456. PMID: [8172042](#)
5. Meissner HC. Economic impact of viral respiratory disease in children. *J Pediatr*. 1994; 124: 17–21.
6. Hakulinen AL, Jarvenpaa AL, Turpeinen M, Sovijarvi A. Diffusing capacity of the lung in school-aged children born very preterm, with and without bronchopulmonary dysplasia. *Pediatr Pulmonol*. 1996; 21: 353–60. doi: [10.1002/\(SICI\)1099-0496\(199606\)21:6<353::AID-PPUL2>3.0.CO;2-M](#) PMID: [8927461](#)
7. Koumbourlis AC, Motoyama EK, Mutich RL, Mallory GB, Walczak SA, Fertal K. Longitudinal follow-up of lung function from childhood to adolescence in prematurely born patients with neonatal chronic lung disease. *Pediatr Pulmonol*. 1996; 21: 28–34. doi: [10.1002/\(SICI\)1099-0496\(199601\)21:1<28::AID-PPUL5>3.0.CO;2-M](#) PMID: [8776263](#)
8. Finkelstein JN, Horowitz S, Sinkin RA, Ryan RM. Cellular and molecular responses to lung injury in relation to induction of tissue repair and fibrosis. *Clin Perinatol*. 1992; 19: 603–20. PMID: [1526074](#)
9. Zimmerman JJ. Bronchoalveolar inflammatory pathophysiology of bronchopulmonary dysplasia. *Clin Perinatol*. 1995; 22: 429–56. PMID: [7671546](#)
10. Zimmerman JJ, Farrell PM. Advances and issues in bronchopulmonary dysplasia. *Curr Probl Pediatr*. 1994; 24: 159–70. PMID: [7915224](#)
11. Kotecha S. Cytokines in chronic lung disease of prematurity. *Eur J Pediatr*. 1996; 155 Suppl 2: 14–7.
12. Groneck P, Speer CP. Inflammatory mediators and bronchopulmonary dysplasia. *Arch Dis Child Fetal Neonatal Ed*. 1995; 73: F1–3. PMID: [7552588](#)
13. Kotecha S, Chan B, Azam N, Silverman M, Shaw RJ. Increase in interleukin-8 and soluble intercellular adhesion molecule-1 in bronchoalveolar lavage fluid from premature infants who develop chronic lung disease. *Arch Dis Child Fetal Neonatal Ed*. 1995; 72: F90–6. PMID: [7712280](#)
14. Kotecha S, Wangoo A, Silverman M, Shaw RJ. Increase in the concentration of transforming growth factor beta-1 in bronchoalveolar lavage fluid before development of chronic lung disease of prematurity. *J Pediatr*. 1996; 128: 464–9. PMID: [8618178](#)
15. Kotecha S, Wilson L, Wangoo A, Silverman M, Shaw RJ. Increase in interleukin (IL)-1 beta and IL-6 in bronchoalveolar lavage fluid obtained from infants with chronic lung disease of prematurity. *Pediatr Res*. 1996; 40: 250–6. doi: [10.1203/00006450-199608000-00010](#) PMID: [8827773](#)
16. Murch SH, Costeloe K, Klein NJ, MacDonald TT. Early production of macrophage inflammatory protein-1 alpha occurs in respiratory distress syndrome and is associated with poor outcome. *Pediatr Res*. 1996; 40: 490–7. doi: [10.1203/00006450-199609000-00020](#) PMID: [8865289](#)
17. Little S, Dean T, Bevin S, Hall M, Ashton M, Church M, et al. Role of elevated plasma soluble ICAM-1 and bronchial lavage fluid IL-8 levels as markers of chronic lung disease in premature infants. *Thorax*. 1995; 50: 1073–9. PMID: [7491556](#)
18. Bagchi A, Viscardi RM, Taciak V, Ensor JE, McCreary KA, Hasday JD. Increased activity of interleukin-6 but not tumor necrosis factor-alpha in lung lavage of premature infants is associated with the development of bronchopulmonary dysplasia. *Pediatr Res*. 1994; 36: 244–52. doi: [10.1203/00006450-199408000-00017](#) PMID: [7970941](#)
19. Groneck P, Gotze-Speer B, Oppermann M, Eiffert H, Speer CP. Association of pulmonary inflammation and increased microvascular permeability during the development of bronchopulmonary dysplasia: a sequential analysis of inflammatory mediators in respiratory fluids of high-risk preterm neonates. *Pediatrics*. 1994; 93: 712–8. PMID: [8165067](#)
20. Thome U, Kossel H, Lipowsky G, Porz F, Furste HO, Genzel-Boroviczeny O, et al. Randomized comparison of high-frequency ventilation with high-rate intermittent positive pressure ventilation in preterm infants with respiratory failure. *J Pediatr*. 1999; 135: 39–46. PMID: [10393602](#)
21. Thome U, Gotze-Speer B, Speer CP, Pohlandt F. Comparison of pulmonary inflammatory mediators in preterm infants treated with intermittent positive pressure ventilation or high frequency oscillatory ventilation. *Pediatr Res*. 1998; 44: 330–7. doi: [10.1203/00006450-199809000-00011](#) PMID: [9727709](#)
22. Ichikawa A, Kuba K, Morita M, Chida S, Tezuka H, Hara H, et al. CXCL10-CXCR3 enhances the development of neutrophil-mediated fulminant lung injury of viral and nonviral origin. *Am J Respir Crit Care Med*. 2013; 187: 65–77. doi: [10.1164/rccm.201203-0508OC](#) PMID: [23144331](#)

23. Lex D, Kuba K, Uhlig S, Imai Y. NPY-Mediated Neuro-Immune Cross Talk In The Pathogenesis Of The Influenza Virus Infection. In: B32. VIRAL INFECTION OF THE AIRWAY: American Thoracic Society; 2014. p. A2738.
24. Rauvala H, Hallman M. Glycolipid accumulation in bronchoalveolar space in adult respiratory distress syndrome. *J Lipid Res.* 1984; 25: 1257–62. PMID: [6520545](#)
25. Kunzmann S, Collins JJP, Yang Y, Uhlig S, Kallapur SG, Speer CP, et al. Antenatal inflammation reduces expression of caveolin-1 and influences multiple signaling pathways in preterm fetal lungs. *Am J Respir Cell Mol Biol.* 2011; 45: 969–76. doi: [10.1165/rcmb.2010-0519OC](#) PMID: [21562314](#)
26. Claus RA, Bunck AC, Bockmeyer CL, Brunkhorst FM, Lösche W, Kinscherf R, et al. Role of increased sphingomyelinase activity in apoptosis and organ failure of patients with severe sepsis. *FASEB J.* 2005; 19: 1719–21. doi: [10.1096/fj.04-2842fje](#) PMID: [16051685](#)
27. Uhlig S, Yang Y. Sphingolipids in acute lung injury. *Handb Exp Pharmacol.* 2013: 227–46.
28. Bismarck P von, Wistädt CG, Klemm K, Winoto-Morbach S, Uhlig U, Schütze S, et al. Improved pulmonary function by acid sphingomyelinase inhibition in a newborn piglet lavage model. *Am J Respir Crit Care Med.* 2008; 177: 1233–41. doi: [10.1164/rccm.200705-752OC](#) PMID: [18310483](#)
29. Preuss S, Omam FD, Scheiermann J, Stadelmann S, Winoto-Morbach S, Bismarck P von, et al. Topical application of phosphatidyl-inositol-3,5-bisphosphate for acute lung injury in neonatal swine. *J Cell Mol Med.* 2012; 16: 2813–26. doi: [10.1111/j.1582-4934.2012.01618.x](#) PMID: [22882773](#)
30. Chan C, Goldkorn T. Ceramide path in human lung cell death. *Am J Respir Cell Mol Biol.* 2000; 22: 460–8. doi: [10.1165/ajrcmb.22.4.3376](#) PMID: [10745027](#)
31. Lavrentiadou SN, Chan C, Kawcak T, Ravid T, Tsaba A, van der Vliet A, et al. Ceramide-mediated apoptosis in lung epithelial cells is regulated by glutathione. *Am J Respir Cell Mol Biol.* 2001; 25: 676–84. doi: [10.1165/ajrcmb.25.6.4321](#) PMID: [11726392](#)
32. Honda K, Kobayashi H, Hataishi R, Hirano S, Fukuyama N, Nakazawa H, et al. Inhaled nitric oxide reduces tyrosine nitration after lipopolysaccharide instillation into lungs of rats. *Am J Respir Crit Care Med.* 1999; 160: 678–88. doi: [10.1164/ajrcm.160.2.9807112](#) PMID: [10430746](#)
33. Kinsella JP, Parker TA, Galan H, Sheridan BC, Halbower AC, Abman SH. Effects of inhaled nitric oxide on pulmonary edema and lung neutrophil accumulation in severe experimental hyaline membrane disease. *Pediatr Res.* 1997; 41: 457–63. doi: [10.1203/00006450-199704000-00002](#) PMID: [9098845](#)
34. Tang J, Markham NE, Lin Y, McMurtry IF, Maxey A, Kinsella JP, et al. Inhaled nitric oxide attenuates pulmonary hypertension and improves lung growth in infant rats after neonatal treatment with a VEGF receptor inhibitor. *Am J Physiol Lung Cell Mol Physiol.* 2004; 287: L344–51. doi: [10.1152/ajplung.00291.2003](#) PMID: [15064225](#)
35. Issa A, Lappalainen U, Kleinman M, Bry K, Hallman M. Inhaled nitric oxide decreases hyperoxia-induced surfactant abnormality in preterm rabbits. *Pediatr Res.* 1999; 45: 247–54. doi: [10.1203/00006450-199902000-00016](#) PMID: [10022598](#)
36. Lin Y, Markham NE, Balasubramaniam V, Tang J, Maxey A, Kinsella JP, et al. Inhaled nitric oxide enhances distal lung growth after exposure to hyperoxia in neonatal rats. *Pediatr Res.* 2005; 58: 22–9. doi: [10.1203/01.PDR.0000163378.94837.3E](#) PMID: [15879297](#)
37. McCurnin DC, Pierce RA, Chang LY, Gibson LL, Osborne-Lawrence S, Yoder BA, et al. Inhaled NO improves early pulmonary function and modifies lung growth and elastin deposition in a baboon model of neonatal chronic lung disease. *Am J Physiol Lung Cell Mol Physiol.* 2005; 288: L450–9. doi: [10.1152/ajplung.00347.2004](#) PMID: [15591412](#)
38. Bland RD, Albertine KH, Carlton DP, MacRitchie AJ. Inhaled nitric oxide effects on lung structure and function in chronically ventilated preterm lambs. *Am J Respir Crit Care Med.* 2005; 172: 899–906. doi: [10.1164/rccm.200503-384OC](#) PMID: [15976381](#)
39. Ballard PL, Gonzales LW, Godinez RI, Godinez MH, Savani RC, McCurnin DC, et al. Surfactant composition and function in a primate model of infant chronic lung disease: effects of inhaled nitric oxide. *Pediatr Res.* 2006; 59: 157–62. doi: [10.1203/01.pdr.0000190664.69081.f1](#) PMID: [16326985](#)
40. Duong-Quy S, Hua-Huy T, Pham H, Tang X, Mercier JC, Baud O, et al. Early inhaled nitric oxide at high dose enhances rat lung development after birth. *Nitric Oxide.* 2014; 38: 8–16. doi: [10.1016/j.niox.2014.02.004](#) PMID: [24566008](#)
41. Rose MJ, Stenger MR, Joshi MS, Welty SE, Bauer JA, Nelin LD. Inhaled nitric oxide decreases leukocyte trafficking in the neonatal mouse lung during exposure to 95% oxygen. *Pediatr Res.* 2010; 67: 244–9. doi: [10.1203/PDR.0b013e3181ca0d93](#) PMID: [19915514](#)
42. Schreiber MD, Gin-Mestan K, Marks JD, Huo D, Lee G, Srisuparp P. Inhaled nitric oxide in premature infants with the respiratory distress syndrome. *N Engl J Med.* 2003; 349: 2099–107. doi: [10.1056/NEJMoa031154](#) PMID: [14645637](#)

43. van Meurs KP, Wright LL, Ehrenkranz RA, Lemons JA, Ball MB, Poole WK, et al. Inhaled nitric oxide for premature infants with severe respiratory failure. *N Engl J Med*. 2005; 353: 13–22. doi: [10.1056/NEJMoa043927](https://doi.org/10.1056/NEJMoa043927) PMID: [16000352](https://pubmed.ncbi.nlm.nih.gov/16000352/)
44. Hascoet JM, Fresson J, Claris O, Hamon I, Lomet J, Liska A, et al. The safety and efficacy of nitric oxide therapy in premature infants. *J Pediatr*. 2005; 146: 318–23. doi: [10.1016/j.jpeds.2004.10.019](https://doi.org/10.1016/j.jpeds.2004.10.019) PMID: [15756211](https://pubmed.ncbi.nlm.nih.gov/15756211/)
45. Mercier J, Hummler H, Durrmeyer X, Sanchez-Luna M, Carnielli V, Field D, et al. Inhaled nitric oxide for prevention of bronchopulmonary dysplasia in premature babies (EUNO): a randomised controlled trial. *Lancet*. 2010; 376: 346–54. doi: [10.1016/S0140-6736\(10\)60664-2](https://doi.org/10.1016/S0140-6736(10)60664-2) PMID: [20655106](https://pubmed.ncbi.nlm.nih.gov/20655106/)
46. Beckman JS, Koppenol WH. Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and ugly. *Am J Physiol*. 1996; 271: C1424–37. PMID: [8944624](https://pubmed.ncbi.nlm.nih.gov/8944624/)
47. Zhu S, Manuel M, Tanaka S, Choe N, Kagan E, Matalon S. Contribution of reactive oxygen and nitrogen species to particulate-induced lung injury. *Environ Health Perspect*. 1998; 106 Suppl 5: 1157–63.
48. Sittipunt C, Steinberg KP, Ruzinski JT, Myles C, Zhu S, Goodman RB, et al. Nitric oxide and nitrotyrosine in the lungs of patients with acute respiratory distress syndrome. *Am J Respir Crit Care Med*. 2001; 163: 503–10. doi: [10.1164/ajrccm.163.2.2004187](https://doi.org/10.1164/ajrccm.163.2.2004187) PMID: [11179131](https://pubmed.ncbi.nlm.nih.gov/11179131/)
49. Zhu S, Ware LB, Geiser T, Matthay MA, Matalon S. Increased levels of nitrate and surfactant protein a nitration in the pulmonary edema fluid of patients with acute lung injury. *Am J Respir Crit Care Med*. 2001; 163: 166–72. doi: [10.1164/ajrccm.163.1.2005068](https://doi.org/10.1164/ajrccm.163.1.2005068) PMID: [11208643](https://pubmed.ncbi.nlm.nih.gov/11208643/)
50. Zhu S, Haddad IY, Matalon S. Nitration of surfactant protein A (SP-A) tyrosine residues results in decreased mannose binding ability. *Arch Biochem Biophys*. 1996; 333: 282–90. doi: [10.1006/abbi.1996.0392](https://doi.org/10.1006/abbi.1996.0392) PMID: [8806782](https://pubmed.ncbi.nlm.nih.gov/8806782/)
51. Rubbo H, Radi R, Anselmi D, Kirk M, Barnes S, Butler J, et al. Nitric oxide reaction with lipid peroxyl radicals spares alpha-tocopherol during lipid peroxidation. Greater oxidant protection from the pair nitric oxide/alpha-tocopherol than alpha-tocopherol/ascorbate. *J Biol Chem*. 2000; 275: 10812–8. PMID: [10753874](https://pubmed.ncbi.nlm.nih.gov/10753874/)
52. O'Donnell VB, Chumley PH, Hogg N, Bloodsworth A, Darley-Usmar VM, Freeman BA. Nitric oxide inhibition of lipid peroxidation: kinetics of reaction with lipid peroxyl radicals and comparison with alpha-tocopherol. *Biochemistry*. 1997; 36: 15216–23. doi: [10.1021/bi971891z](https://doi.org/10.1021/bi971891z) PMID: [9398249](https://pubmed.ncbi.nlm.nih.gov/9398249/)
53. Walsh MC, Wilson-Costello D, Zadell A, Newman N, Fanaroff A. Safety, reliability, and validity of a physiologic definition of bronchopulmonary dysplasia. *J Perinatol*. 2003; 23: 451–6. doi: [10.1038/sj.jp.7210963](https://doi.org/10.1038/sj.jp.7210963) PMID: [13679930](https://pubmed.ncbi.nlm.nih.gov/13679930/)
54. Walsh MC, Yao Q, Gettner P, Hale E, Collins M, Hensman A, et al. Impact of a physiologic definition on bronchopulmonary dysplasia rates. *Pediatrics*. 2004; 114: 1305–11. doi: [10.1542/peds.2004-0204](https://doi.org/10.1542/peds.2004-0204) PMID: [15520112](https://pubmed.ncbi.nlm.nih.gov/15520112/)
55. Wrigge H, Uhlig U, Zinserling J, Behrends-Callsen E, Ottersbach G, Fischer M, et al. The effects of different ventilatory settings on pulmonary and systemic inflammatory responses during major surgery. *Anesth Analg*. 2004; 98: 775–81. PMID: [14980936](https://pubmed.ncbi.nlm.nih.gov/14980936/)
56. Goggel R, Winoto-Morbach S, Vielhaber G, Imai Y, Lindner K, Brade L, et al. PAF-mediated pulmonary edema: a new role for acid sphingomyelinase and ceramide. *Nat Med*. 2004; 10: 155–60. doi: [10.1038/nm977](https://doi.org/10.1038/nm977) PMID: [14704790](https://pubmed.ncbi.nlm.nih.gov/14704790/)
57. Blic J de, Midulla F, Barbato A, Clement A, Dab I, Eber E, et al. Bronchoalveolar lavage in children. ERS Task Force on bronchoalveolar lavage in children. European Respiratory Society. *Eur Respir J*. 2000; 15: 217–31. PMID: [10678650](https://pubmed.ncbi.nlm.nih.gov/10678650/)
58. Haslam PL, Baughman RP. Report of ERS Task Force: guidelines for measurement of acellular components and standardization of BAL. *Eur Respir J*. 1999; 14: 245–8. PMID: [10515395](https://pubmed.ncbi.nlm.nih.gov/10515395/)
59. Ambalavanan N, Novak ZE. Peptide growth factors in tracheal aspirates of mechanically ventilated preterm neonates. *Pediatr Res*. 2003; 53: 240–4. doi: [10.1203/01.PDR.0000047656.17766.39](https://doi.org/10.1203/01.PDR.0000047656.17766.39) PMID: [12538781](https://pubmed.ncbi.nlm.nih.gov/12538781/)
60. Jonsson B, Li YH, Noack G, Brauner A, Tullus K. Downregulatory cytokines in tracheobronchial aspirate fluid from infants with chronic lung disease of prematurity. *Acta Paediatr*. 2000; 89: 1375–80. PMID: [11106053](https://pubmed.ncbi.nlm.nih.gov/11106053/)
61. Thompson A, Bhandari V. Pulmonary Biomarkers of Bronchopulmonary Dysplasia. *Biomark Insights*. 2008; 3: 361–73. PMID: [19430584](https://pubmed.ncbi.nlm.nih.gov/19430584/)
62. Truog WE, Ballard PL, Norberg M, Golombek S, Savani RC, Merrill JD, et al. Inflammatory markers and mediators in tracheal fluid of premature infants treated with inhaled nitric oxide. *Pediatrics*. 2007; 119: 670–8. doi: [10.1542/peds.2006-2683](https://doi.org/10.1542/peds.2006-2683) PMID: [17403837](https://pubmed.ncbi.nlm.nih.gov/17403837/)

63. Dezfulian C, Raat N, Shiva S, Gladwin MT. Role of the anion nitrite in ischemia-reperfusion cytoprotection and therapeutics. *Cardiovasc Res*. 2007; 75: 327–38. doi: [10.1016/j.cardiores.2007.05.001](https://doi.org/10.1016/j.cardiores.2007.05.001) PMID: [17568573](https://pubmed.ncbi.nlm.nih.gov/17568573/)
64. Posencheg MA, Gow AJ, Truog WE, Ballard RA, Cnaan A, Golombek SG, et al. Inhaled nitric oxide in premature infants: effect on tracheal aspirate and plasma nitric oxide metabolites. *J Perinatol*. 2010; 30: 275–80. doi: [10.1038/jp.2009.155](https://doi.org/10.1038/jp.2009.155) PMID: [19812581](https://pubmed.ncbi.nlm.nih.gov/19812581/)
65. Banks BA, Ischiropoulos H, McClelland M, Ballard PL, Ballard RA. Plasma 3-nitrotyrosine is elevated in premature infants who develop bronchopulmonary dysplasia. *Pediatrics*. 1998; 101: 870–4. PMID: [9565417](https://pubmed.ncbi.nlm.nih.gov/9565417/)
66. Lorch SA, Banks BA, Christie J, Merrill JD, Althaus J, Schmidt K, et al. Plasma 3-nitrotyrosine and outcome in neonates with severe bronchopulmonary dysplasia after inhaled nitric oxide. *Free Radic Biol Med*. 2003; 34: 1146–52. PMID: [12706495](https://pubmed.ncbi.nlm.nih.gov/12706495/)
67. Ballard PL, Truog WE, Merrill JD, Gow A, Posencheg M, Golombek SG, et al. Plasma biomarkers of oxidative stress: relationship to lung disease and inhaled nitric oxide therapy in premature infants. *Pediatrics*. 2008; 121: 555–61. doi: [10.1542/peds.2007-2479](https://doi.org/10.1542/peds.2007-2479) PMID: [18310205](https://pubmed.ncbi.nlm.nih.gov/18310205/)
68. Ballard RA, Truog WE, Cnaan A, Martin RJ, Ballard PL, Merrill JD, et al. Inhaled nitric oxide in preterm infants undergoing mechanical ventilation. *N Engl J Med*. 2006; 355: 343–53. doi: [10.1056/NEJMoa061088](https://doi.org/10.1056/NEJMoa061088) PMID: [16870913](https://pubmed.ncbi.nlm.nih.gov/16870913/)
69. Bartram U, Speer CP. The role of transforming growth factor beta in lung development and disease. *Chest*. 2004; 125: 754–65. PMID: [14769761](https://pubmed.ncbi.nlm.nih.gov/14769761/)
70. Beers MF, Solarin KO, Guttentag SH, Rosenbloom J, Kormilli A, Gonzales LW, et al. TGF-beta1 inhibits surfactant component expression and epithelial cell maturation in cultured human fetal lung. *Am J Physiol*. 1998; 275: L950–60. PMID: [9815113](https://pubmed.ncbi.nlm.nih.gov/9815113/)
71. Ballard PL, Merrill JD, Truog WE, Godinez RI, Godinez MH, McDevitt TM, et al. Surfactant function and composition in premature infants treated with inhaled nitric oxide. *Pediatrics*. 2007; 120: 346–53. doi: [10.1542/peds.2007-0095](https://doi.org/10.1542/peds.2007-0095) PMID: [17671061](https://pubmed.ncbi.nlm.nih.gov/17671061/)
72. Hashimoto S, Gon Y, Takeshita I, Matsumoto K, Maruoka S, Horie T. Transforming growth Factor-beta1 induces phenotypic modulation of human lung fibroblasts to myofibroblast through a c-Jun-NH2-terminal kinase-dependent pathway. *Am J Respir Crit Care Med*. 2001; 163: 152–7. doi: [10.1164/ajrccm.163.1.2005069](https://doi.org/10.1164/ajrccm.163.1.2005069) PMID: [11208641](https://pubmed.ncbi.nlm.nih.gov/11208641/)
73. Kasai H, Allen JT, Mason RM, Kamimura T, Zhang Z. TGF-beta1 induces human alveolar epithelial to mesenchymal cell transition (EMT). *Respir Res*. 2005; 6: 56. doi: [10.1186/1465-9921-6-56](https://doi.org/10.1186/1465-9921-6-56) PMID: [15946381](https://pubmed.ncbi.nlm.nih.gov/15946381/)
74. Xu L, Xiang X, Ji X, Wang W, Luo M, Luo S, et al. Effects and mechanism of dehydroepiandrosterone on epithelial-mesenchymal transition in bronchial epithelial cells. *Exp Lung Res*. 2014; 40: 211–21. doi: [10.3109/01902148.2013.879966](https://doi.org/10.3109/01902148.2013.879966) PMID: [24784499](https://pubmed.ncbi.nlm.nih.gov/24784499/)
75. Vicencio AG, Lee CG, Cho SJ, Eickelberg O, Chuu Y, Haddad GG, et al. Conditional overexpression of bioactive transforming growth factor-beta1 in neonatal mouse lung: a new model for bronchopulmonary dysplasia? *Am J Respir Cell Mol Biol*. 2004; 31: 650–6. doi: [10.1165/rcmb.2004-0092OC](https://doi.org/10.1165/rcmb.2004-0092OC) PMID: [15333328](https://pubmed.ncbi.nlm.nih.gov/15333328/)
76. Vyas-Read S, Shaul PW, Yuhanna IS, Willis BC. Nitric oxide attenuates epithelial-mesenchymal transition in alveolar epithelial cells. *Am J Physiol Lung Cell Mol Physiol*. 2007; 293: L212–21. doi: [10.1152/ajplung.00475.2006](https://doi.org/10.1152/ajplung.00475.2006) PMID: [17496059](https://pubmed.ncbi.nlm.nih.gov/17496059/)
77. ter Horst SAJ, Walthers FJ, Poorthuis BJHM, Hiemstra PS, Wagenaar GTM. Inhaled nitric oxide attenuates pulmonary inflammation and fibrin deposition and prolongs survival in neonatal hyperoxic lung injury. *Am J Physiol Lung Cell Mol Physiol*. 2007; 293: L35–44. doi: [10.1152/ajplung.00381.2006](https://doi.org/10.1152/ajplung.00381.2006) PMID: [17384081](https://pubmed.ncbi.nlm.nih.gov/17384081/)
78. Roux J, Carles M, Koh H, Goolaerts A, Ganter MT, Chesebro BB, et al. Transforming growth factor beta1 inhibits cystic fibrosis transmembrane conductance regulator-dependent cAMP-stimulated alveolar epithelial fluid transport via a phosphatidylinositol 3-kinase-dependent mechanism. *J Biol Chem*. 2010; 285: 4278–90. doi: [10.1074/jbc.M109.036731](https://doi.org/10.1074/jbc.M109.036731) PMID: [19996317](https://pubmed.ncbi.nlm.nih.gov/19996317/)
79. Wagener BM, Roux J, Carles M, Pittet J. Synergistic Inhibition of beta2-adrenergic Receptor-mediated Alveolar Epithelial Fluid Transport by Interleukin-8 and Transforming Growth Factor-beta. *Anesthesiology*. 2015; 122(5): 1084–92. doi: [10.1097/ALN.0000000000000595](https://doi.org/10.1097/ALN.0000000000000595) PMID: [25591042](https://pubmed.ncbi.nlm.nih.gov/25591042/)
80. Aghai ZH, Saslow JG, Mody K, Eydelman R, Bhat V, Stahl G, et al. IFN-gamma and IP-10 in tracheal aspirates from premature infants: relationship with bronchopulmonary dysplasia. *Pediatr Pulmonol*. 2013; 48: 8–13. doi: [10.1002/ppul.22540](https://doi.org/10.1002/ppul.22540) PMID: [22431160](https://pubmed.ncbi.nlm.nih.gov/22431160/)
81. Hikino S, Ohga S, Kinjo T, Kusuda T, Ochiai M, Inoue H, et al. Tracheal aspirate gene expression in preterm newborns and development of bronchopulmonary dysplasia. *Pediatr Int*. 2012; 54: 208–14. doi: [10.1111/j.1442-200X.2011.03510.x](https://doi.org/10.1111/j.1442-200X.2011.03510.x) PMID: [22066648](https://pubmed.ncbi.nlm.nih.gov/22066648/)

82. Harijith A, Choo-Wing R, Cataltepe S, Yasumatsu R, Aghai ZH, Janer J, et al. A role for matrix metalloproteinase 9 in IFN γ -mediated injury in developing lungs: relevance to bronchopulmonary dysplasia. *Am J Respir Cell Mol Biol*. 2011; 44: 621–30. doi: [10.1165/rcmb.2010-0058OC](https://doi.org/10.1165/rcmb.2010-0058OC) PMID: [21216975](https://pubmed.ncbi.nlm.nih.gov/21216975/)
83. Munshi UK, Niu JO, Siddiq MM, Parton LA. Elevation of interleukin-8 and interleukin-6 precedes the influx of neutrophils in tracheal aspirates from preterm infants who develop bronchopulmonary dysplasia. *Pediatr Pulmonol*. 1997; 24: 331–6. PMID: [9407566](https://pubmed.ncbi.nlm.nih.gov/9407566/)
84. McCole JM, McIntosh N. Interleukin-8 in bronchoalveolar lavage samples as predictor of chronic lung disease in premature infants. *Lancet*. 1994; 343: 729.
85. Niu JO, Munshi UK, Siddiq MM, Parton LA. Early increase in endothelin-1 in tracheal aspirates of preterm infants: correlation with bronchopulmonary dysplasia. *J Pediatr*. 1998; 132: 965–70. PMID: [9627587](https://pubmed.ncbi.nlm.nih.gov/9627587/)
86. Kwong KY, Jones CA, Cayabyab R, Lecart C, Stotts CL, Randhawa J, et al. Differential regulation of IL-8 by IL-1 β and TNF α in hyaline membrane disease. *J Clin Immunol*. 1998; 18: 71–80. PMID: [9475356](https://pubmed.ncbi.nlm.nih.gov/9475356/)
87. El Kebir D, Hubert B, Taha R, Troncy E, Wang T, Gauvin D, et al. Effects of inhaled nitric oxide on inflammation and apoptosis after cardiopulmonary bypass. *Chest*. 2005; 128: 2910–7. doi: [10.1378/chest.128.4.2910](https://doi.org/10.1378/chest.128.4.2910) PMID: [16236968](https://pubmed.ncbi.nlm.nih.gov/16236968/)
88. Chollet-Martin S, Gatecel C, Kermarrec N, Gougerot-Pocidal MA, Payen DM. Alveolar neutrophil functions and cytokine levels in patients with the adult respiratory distress syndrome during nitric oxide inhalation. *Am J Respir Crit Care Med*. 1996; 153: 985–90. doi: [10.1164/ajrccm.153.3.8630584](https://doi.org/10.1164/ajrccm.153.3.8630584) PMID: [8630584](https://pubmed.ncbi.nlm.nih.gov/8630584/)
89. Knox CD, Kam P-J de, Azer K, Wong P, Ederveen AG, Shevell D, et al. Discovery and Clinical Evaluation of MK-8150, A Novel Nitric Oxide Donor With a Unique Mechanism of Nitric Oxide Release. *J Am Heart Assoc*. 2016; 5:e003493. doi: [10.1161/JAHA.116.003493](https://doi.org/10.1161/JAHA.116.003493) PMID: [27561272](https://pubmed.ncbi.nlm.nih.gov/27561272/)
90. Yang Y, Uhlig S. The role of sphingolipids in respiratory disease. *Ther Adv Respir Dis*. 2011; 5: 325–44. doi: [10.1177/1753465811406772](https://doi.org/10.1177/1753465811406772) PMID: [21900155](https://pubmed.ncbi.nlm.nih.gov/21900155/)
91. Cornell TT, Hinkovska-Galcheva V, Sun L, Cai Q, Hershenson MB, Vanway S, et al. Ceramide-dependent PP2A regulation of TNF α -induced IL-8 production in respiratory epithelial cells. *Am J Physiol Lung Cell Mol Physiol*. 2009; 296: L849–56. doi: [10.1152/ajplung.90516.2008](https://doi.org/10.1152/ajplung.90516.2008) PMID: [19286927](https://pubmed.ncbi.nlm.nih.gov/19286927/)
92. Kolliputi N, Galam L, Parthasarathy PT, Tipparaju SM, Lockey RF. NALP-3 inflammasome silencing attenuates ceramide-induced transepithelial permeability. *J Cell Physiol*. 2012; 227: 3310–6. doi: [10.1002/jcp.24026](https://doi.org/10.1002/jcp.24026) PMID: [22169929](https://pubmed.ncbi.nlm.nih.gov/22169929/)
93. Kumar P. Use of inhaled nitric oxide in preterm infants. *Pediatrics*. 2014; 133: 164–70. doi: [10.1542/peds.2013-3444](https://doi.org/10.1542/peds.2013-3444) PMID: [24379225](https://pubmed.ncbi.nlm.nih.gov/24379225/)
94. Cole FS, Alleyne C, Barks, John D E, Boyle RJ, Carroll JL, Dokken D, et al. NIH Consensus Development Conference statement: inhaled nitric-oxide therapy for premature infants. *Pediatrics*. 2011; 127: 363–9. doi: [10.1542/peds.2010-3507](https://doi.org/10.1542/peds.2010-3507) PMID: [21220405](https://pubmed.ncbi.nlm.nih.gov/21220405/)
95. Johnson AH, Peacock JL, Greenough A, Marlow N, Limb ES, Marston L, et al. High-frequency oscillatory ventilation for the prevention of chronic lung disease of prematurity. *N Engl J Med*. 2002; 347: 633–42. doi: [10.1056/NEJMoa020432](https://doi.org/10.1056/NEJMoa020432) PMID: [12200550](https://pubmed.ncbi.nlm.nih.gov/12200550/)
96. Zivanovic S, Peacock J, Alcazar-Paris M, Lo JW, Lunt A, Marlow N, et al. Late outcomes of a randomized trial of high-frequency oscillation in neonates. *N Engl J Med*. 2014; 370: 1121–30. doi: [10.1056/NEJMoa1309220](https://doi.org/10.1056/NEJMoa1309220) PMID: [24645944](https://pubmed.ncbi.nlm.nih.gov/24645944/)